

Peptide search algorithm by selecting and rescoring reliable peaks for MSⁿ (n > 1) spectra

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Overview:

A database search algorithm using MSⁿ (n > 1) spectra obtained by MALDI-IT/MS.

1. Introduction:

Ion trap mass spectrometers have been widely used to analyze the detailed structure of proteins because MSⁿ (n > 1) spectra can be obtained by repeating the isolation and dissociation of precursor ions. MSⁿ (n > 1) spectra, especially where n > 2, include important information and have high potential for analyzing the post-translational modification of proteins. However, conventional software doesn't have sufficient ability to utilize this information in MSⁿ (n > 2) because it does not have any functions to compare and combine the results of MS² with MSⁿ (n > 2). In this study, we propose a novel algorithm for database search which provides more reliable results by using not only MS² but also MSⁿ (n > 2).

2. Method

2.1 Method Overview:

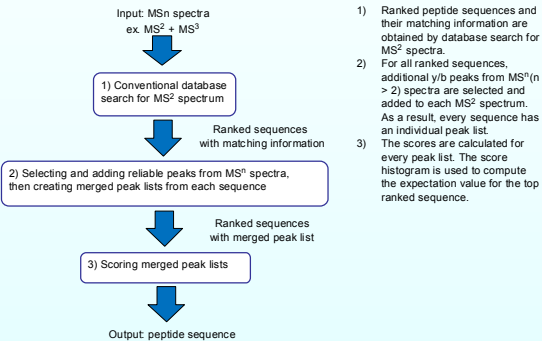


Fig1. Flowchart of this algorithm. In 1), the matching information includes peaks in each spectrum corresponding to theoretical fragments of y/b ion series.

Reference:

[1] Bioinformatics, 20, 1466-1467 (2004), Tandem: matching proteins with tandem mass spectra

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2.2 Example of this algorithm:

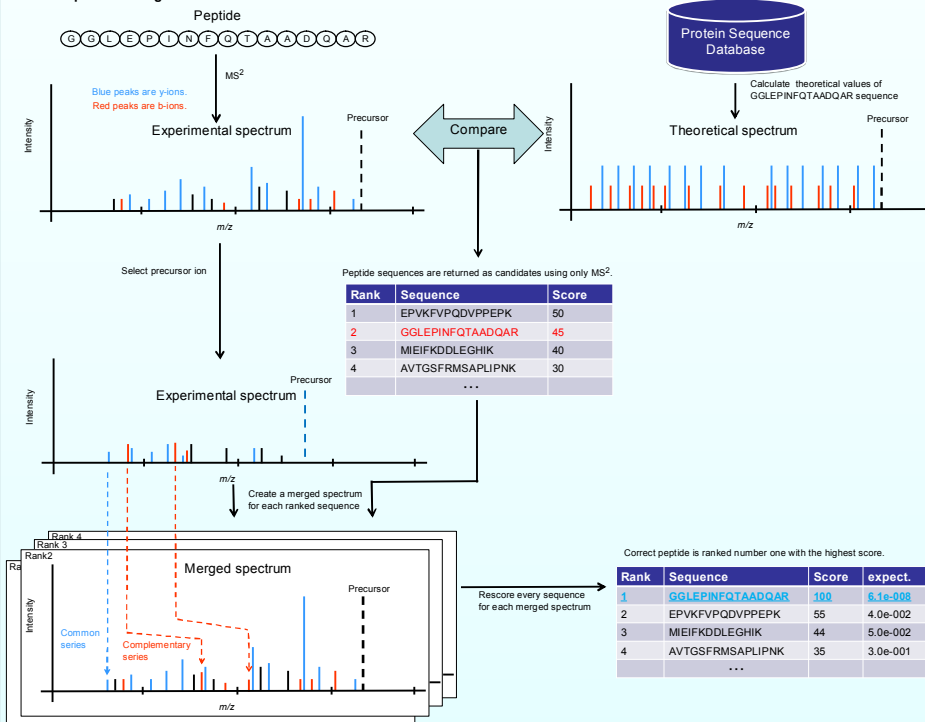


Fig2. Example of this algorithm for the peptide sequence GGLEPINFQTAADQAR. Conventional database search is shown above. After comparison of MS² experimental spectra with theoretical spectra, merged spectra for each sequence are created by adding common ion series and complementary ion series from MS² to MS³. Common ion series have the same masses in both MS² and MS³. Complementary ion series have shifted masses from MS² by the difference between precursor masses in MS² and MS³. When the precursor is a b-ion, y-ions in MS³ are shifted further by the mass of H₂O.

3. Result:

We applied this algorithm to MS² and MS³ spectra of Trypsin-digested proteins.

Dataset summary:

1) Instrument: AXIMA QIT (Shimadzu/KRATOS, UK), Peak detection software: Mascot distiller (Matrix Science), Sample: Trypsin-digested Bovine Serum Albumin, Lysozyme C, Ovalbumin and Glyceraldehyde-3-phosphate dehydrogenase.

2) The ions with the highest intensity or the second highest intensity in MS² were selected as precursors for MS³.

Implementation summary:

1) We implemented this algorithm using X!Tandem¹¹.

2) We used X!Tandem native scoring as the scoring function.

Table 1. MS² column shows correct peptide sequences and expect. (y/b) column shows expectation values and y/b ion count returned by X!Tandem using only MS². MS³ column shows precursor ions and merged expect. value(y/b) column shows the result after applying this algorithm.

Sample	MS ²	expect. (y/b)	MS ³	merged expect. (y/b)
BSA	LVNELTEFAK	3.7e-003(8/5)	y5(LVNELTEFAK)	1.2e-003(7/5)
	LYLEIAR	4.0e+000(4/4)	y3(LYLEIAR)	8.3e-002(5/4)
	RHPYFAPELLYANK	5.6e-005(5/9)	b9(RHPYFAPELLYANK)	6.6e-008(6/10)
	DAIPENLPLTADFADKDVCK	4.5e-009(12/7)	y21(DAIPENLPLTADFADKDVCK)	1.6e-010(12/10)
	LBEYGFQNALVR	4.3e-004(8/4)	y5(LBEYGFQNALVR)	2.7e-008(11/5)
	KVPQVSTPTLVEVSR	9.8e-007(8/7)	b12(KVPQVSTPTLVEVSR)	2.7e-008(11/8)
RPCFSALTPDETVPK	2.1e-003(3/7)	b10(RPCFSALTPDETVPK)	1.4e-008(5/7)	
		b11(RPCFSALTPDETVPK)	1.2e-002(3/7) (*)	
		y14(RPCFSALTPDETVPK)	7.9e-007(10/7)	
LysC	IVSDGNMNAWAVR	5.5e-001(5/1)	y11(IVSDGNMNAWAVR)	6.6e-002(7/1)
	NTDGDSTDYGLQINSR	3.6e-001(6/0)	y13(NTDGDSTDYGLQINSR)	2.3e-003(9/0)
	NLNCNIPCSALLSSDITASVNCVK	2.5e-006(12/3)	y9(NLNCNIPCSALLSSDITASVNCVK)	2.0e-008(11/5)
OVAL	FESNFNTQATNR	2.6e-004(10/3)	y10(FESNFNTQATNR)	6.1e-008(11/8)
	AFKDEDTQAMPFR	7.8e-004(9/2)	y7(AFKDEDTQAMPFR)	2.5e-004(9/4)
	ISQAVFAHAHEAEAR	3.8e-010(9/11)	y8(ISQAVFAHAHEAEAR)	5.9e-012(13/11)
G3P	GGLEPINFQTAADQAR	2.5e-004(10/3)	y12(GGLEPINFQTAADQAR)	8.1e-008(11/8)
	LTEWTSNNMEER	3.8e-005(9/4)	y10(LTEWTSNNMEER)	5.8e-005(10/4) (*)
	LISWYDNEFGYSNR	2.4e-001(7/1)	y8(LISWYDNEFGYSNR)	6.7e-002(7/3)

In most results, additional peaks of false peptides were few and the scores of false peptides remained low. When testing this algorithm, the differences between the scores of true peptides and false peptides were large.

Although, in a few results (indicated by *), expectation values became worse, because the additional peaks were very few. So only overlapping peaks were detected in both MS² and MS³ spectra in these results; overlapping peaks cannot improve scores and expectation values.

4. Discussion:

There are two outstanding problems.

- How to determine the intensities of additional peaks.
 - How to incorporate overlapping peaks in MS² and MS³ spectra in the score calculation.
- It is difficult to determine the intensities of additional peaks. The score may not be improved, if the intensity of an additional peak is weak. However, false peptides may have the highest scores, if a noise peak is selected as a precursor and intensities of additional peaks are very high.

When testing this algorithm, many overlapping peaks in MS² and MS³ were detected, though they are not reflected in the score. I think they should contribute to the confidence level of peptide identification.

5. Conclusion:

Applying the proposed database search algorithm using MSⁿ spectra

- Peptide sequences could be identified with higher confidence.
- MSⁿ spectra are effectively incorporated in the score calculation.